











Scope of the Presentation



- New wave of plant breeding innovations and their potential impact in the future;
- Overview of the current global regulatory frameworks



1. Advanced genome/transcriptome sequencing or genotyping tools (e.g., NGS systems like Illumina, Pacbio, Nanopore)



MiSeq



NextSeq 500



AVITI



NovaSeq 6000

Nanopore Sequencing

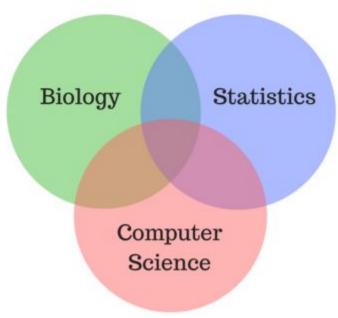


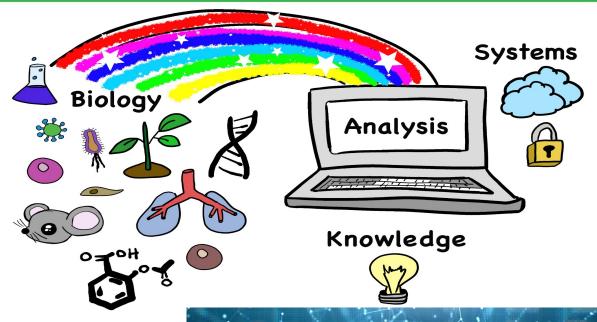


2. Advanced data analytical tools (e.g., bioinformatics, AI)



Bioinformatics

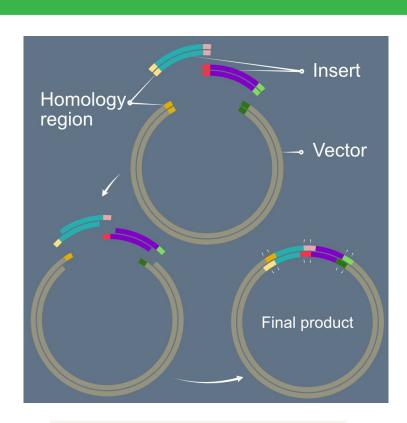






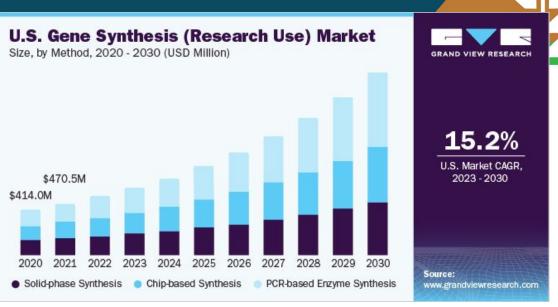


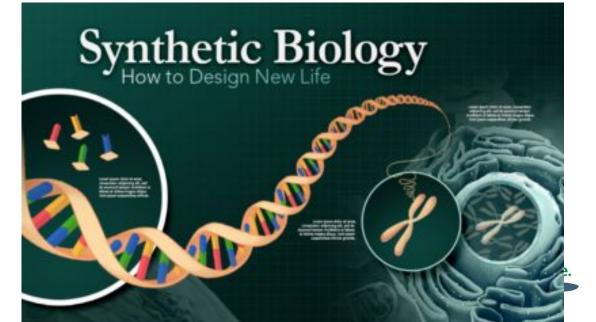
3. Advanced DNA synthesis technologies (up to genome level)



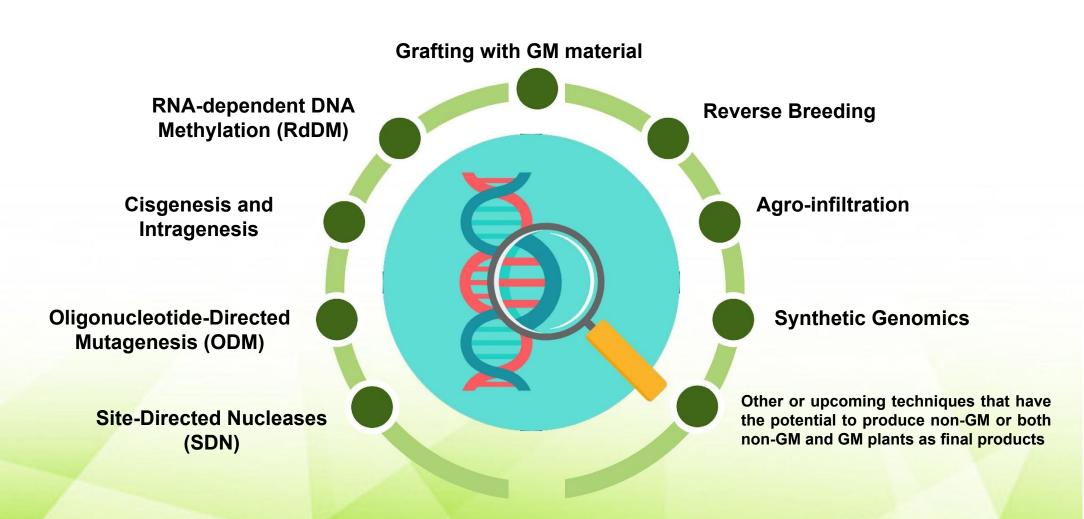
Gibson Assembly

a.k.a Gibson DNA assembly or Gibson cloning





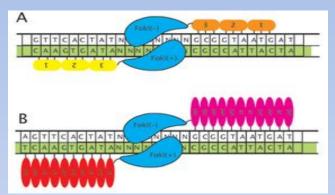
4. Plant Breeding Innovations (including Precision Breeding)



Site-Directed Nucleases or SDN (e.g., ZFN, TALEN, CRISPR-Cas)

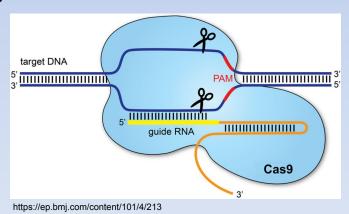
ZFN

TALEN



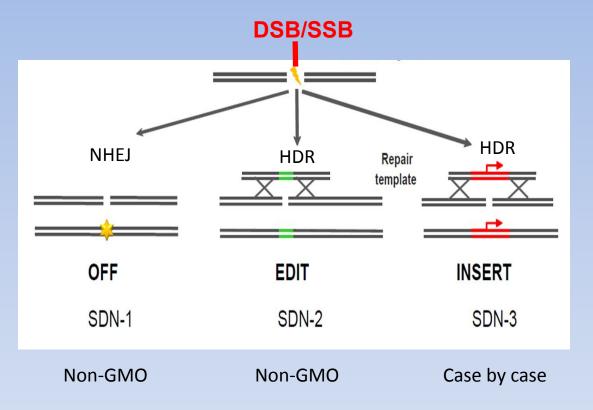
https://www.addgene.org/talen/guide/

CRISPR-Cas9



- DNA-cutting enzyme (nuclease)
 that is directed to cleave the DNA
 at a predetermined location as
 guided by a specific DNA binding
 component, producing a DSB or
 SSB.
- USE: Introduction of a targeted mutation (e.g., insertion or deletion) in a precise location (in the DSB or SSB).

Repair of the double-strand break (DSB)

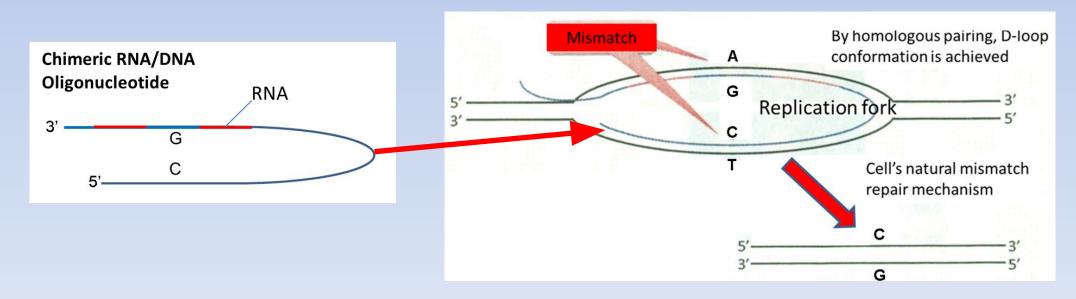


- DSB repair method and nature of insert determine if product is GMO or not
- SDN1 no insertion, no repair template; non-GMO*
- SDN2 insertion of 19 bp and below; non-GMO
- SDN3 insertion of 20 bp and above; case-by-case

^{*}Need to breed out any vector sequences

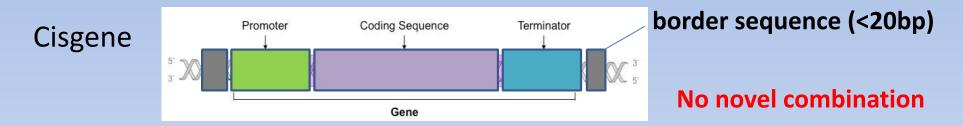
2. Oligonucleotide-directed mutagenesis (ODM)

- Uses customized single-stranded oligonucleotides to interfere with DNA replication at the replication fork within the target gene, resulting in the introduction of desired bases (hence, mutation) into the DNA.
- USE: Introduction of a non-random mutation (e.g., base change) in a precise location to which the designed oligonucleotide is complementary.
- Non-GMO as product

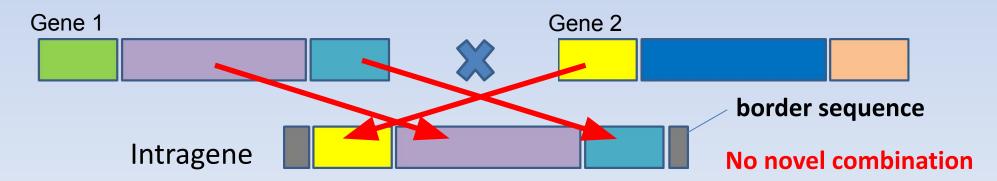


3. Cisgenesis and Intragenesis

 Cisgenesis: genetic transformation of a recipient plant with a natural gene (allele) from a crossable (sexually compatible) plant.

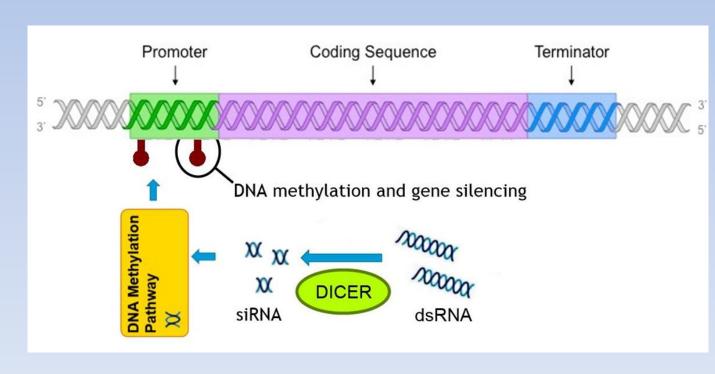


 Intragenesis: Introduces an altered gene (different choice of promoters, exons and introns) from the same or cross-compatible species



4. RNA-dependent DNA methylation (RdDM)

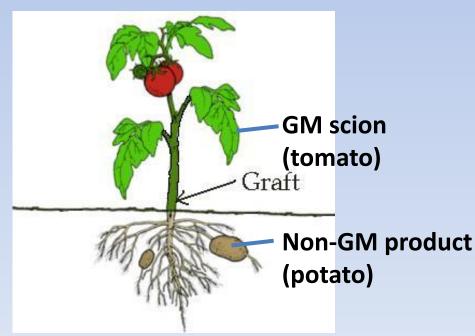
- Knocks down (silences) target genes through RNAi, resulting in the attachment of a -CH3 to their promoter sequence (epigenetic control)
- Methylation does not change the DNA sequence
- Null segregants as final products (non-GMOs)



5. Grafting with GM Material

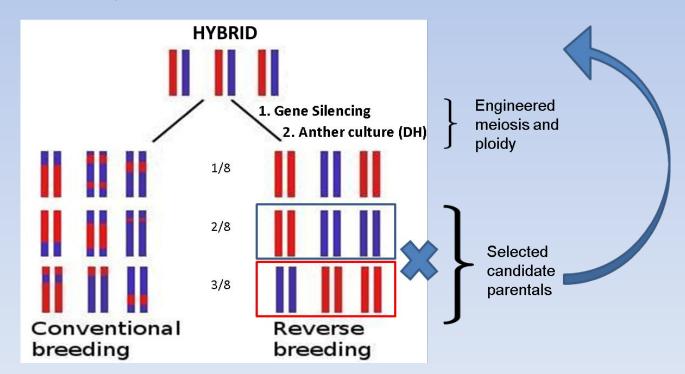
- Fusing together a rootstock and scion, either one being a GMO
- Produces non-GMO products from the non-GMO scion or root stock
- Essential molecules are translocated from the transgenic part but not DNA





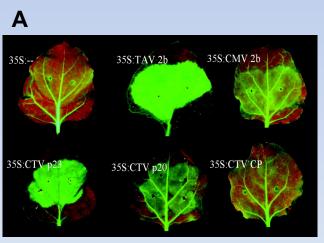
6. Reverse breeding

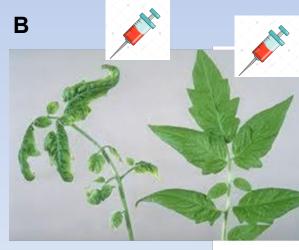
- Prevents "crossing over" in the hybrid (of unknown parentage) during meiosis through RNAi
- Produces homozygous candidate parentals (null segregants) as final products (non-GMO)

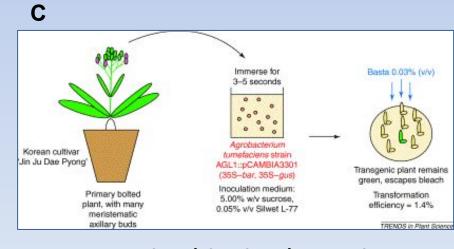


7. Agro-infiltration – Agrobacterium tumefaciens-mediated

- Non-germline (Agroinoculation): Test a construct (transient expression)
 (A) or to express virus particles to screen for resistant plants (B), without affecting the germline tissues of the host
- Germline (Floral dip): Insertion of a gene through the floral parts, immature embryo or meristems that will give rise to seeds (C)





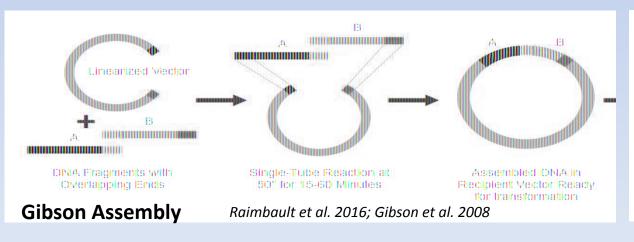


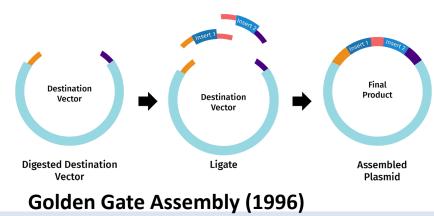
A. Non-germline – no novel combination

B. Germline (Floral Dip) – case by case

8. Synthetic Genomics (overlaps with Synthetic Biology)

- Engineering and manipulation of an organism's genetic material on the scale of the whole genome, based on technologies to design and chemically synthesize pieces of DNA (e.g., Gibson and Golden Gate Assembly) and to assemble them to long, chromosome-sized fragments (Konig et al., 2013)
- As an NBT, it can create a non-GMO by faithfully duplicating the genome
- Can also create a GMO





IRRI leads the way in rice gene editing in PH

> Plant Cell Rep. 2017 May;36(5):745-757. doi: 10.1007/s00299-017-2118-z. Epub 2017 Mar 27.

CRISPR-Cas9 and CRISPR-Cpf1 mediated targeting of a stomatal developmental gene EPFL9 in rice

Xiaojia Yin ¹, Akshaya K Biswal ^{1 2}, Jacqueline Dionora ¹, Kristel M Perdigon ¹, Christian P Balahadia ¹, Shamik Mazumdar ¹, Caspar Chater ³ ⁴, Hsiang-Chun Lin ¹, Robert A Coe ¹, Tobias Kretzschmar ¹, Julie E Gray ³, Paul W Quick ¹, Anindya Bandyopadhyay ⁶

> Plant Biotechnol J. 2018 Nov;16(11):1918-1927. doi: 10.1111/pbi.12927. Epub 2018 Apr 30.

Novel alleles of rice eIF4G generated by CRISPR/Cas9-targeted mutagenesis confer resistance to Rice tungro spherical virus

Anca Macovei ¹, Neah R Sevilla ¹, Christian Cantos ¹, Gilda B Jonson ¹, Inez Slamet-Loedin ¹, Tomáš Čermák ², Daniel F Voytas ², Il-Ryong Choi ¹, Prabhjit Chadha-Mohanty ¹

Broad-spectrum resistance to bacterial blight in rice using genome editing

Ricardo Oliva , Chonghui Ji, Genelou Atienza-Grande, José C. Huguet-Tapia, Alvaro Perez-Quintero, Ting Li, Joon-Seob Eom, Chenhao Li, Hanna Nguyen, Bo Liu, Florence Auguy, Coline Sciallano, Van T. Luu, Gerbert S. Dossa, Sébastien Cunnac, Sarah M. Schmidt, Inez H. Slamet-Loedin, Casiana Vera Cruz, Boris Szurek, Wolf B. Frommer A. Frank F. White & Bing Yang

Nature Biotechnology 37, 1344–1350 (2019) Cite this article

2017

2018

2019









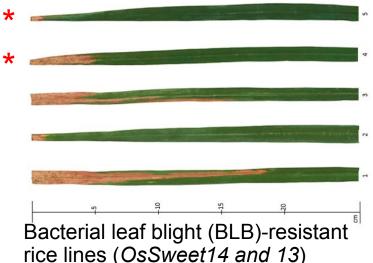
Capacitating PhilRice for genome editing research







RTSV-resistant rice lines (*eIF4g*)

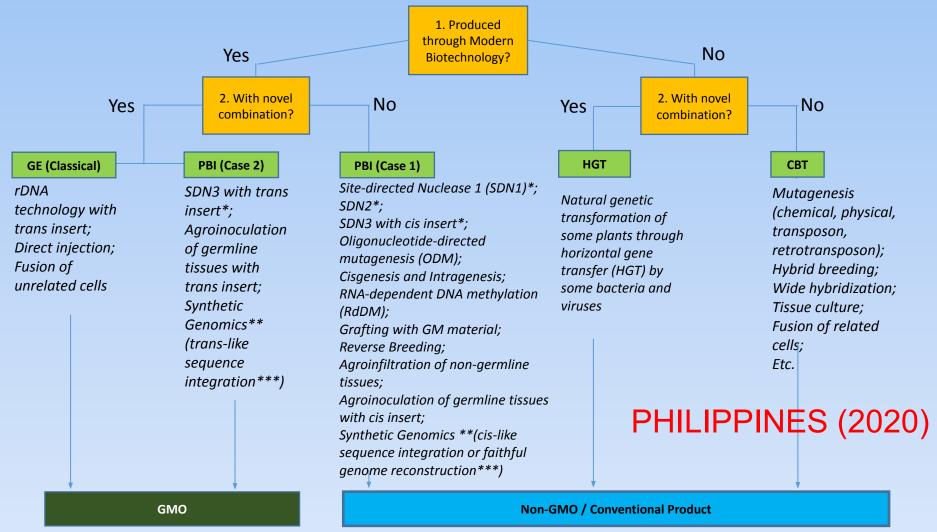


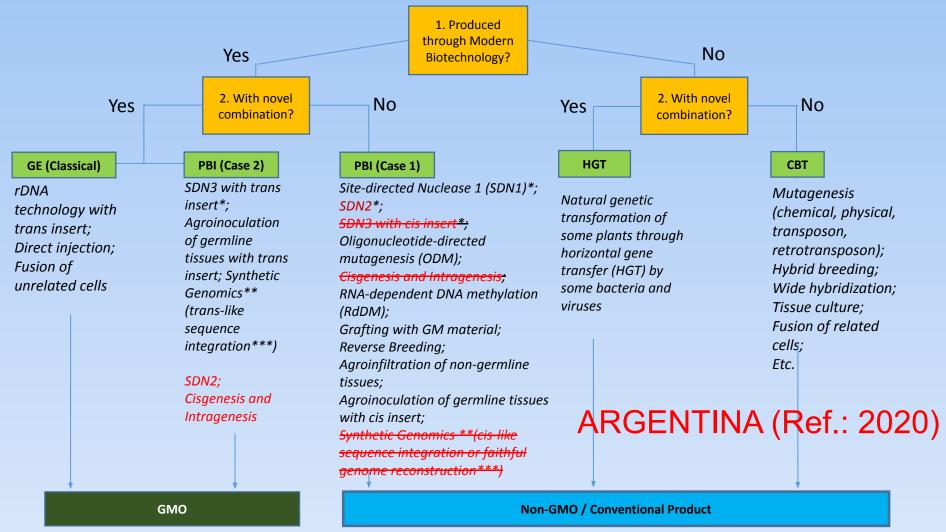
rice lines (OsSweet14 and 13)

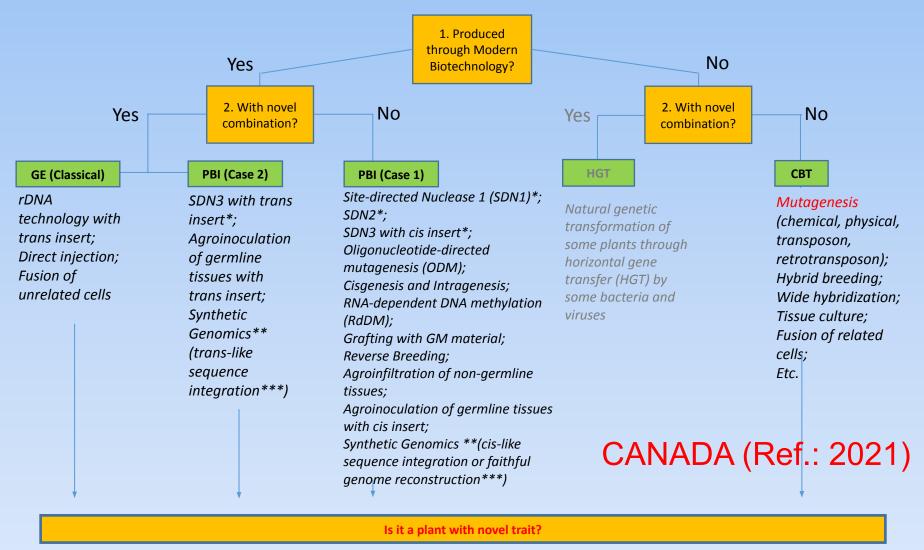


Overview of the current global regulatory frameworks on PBIs

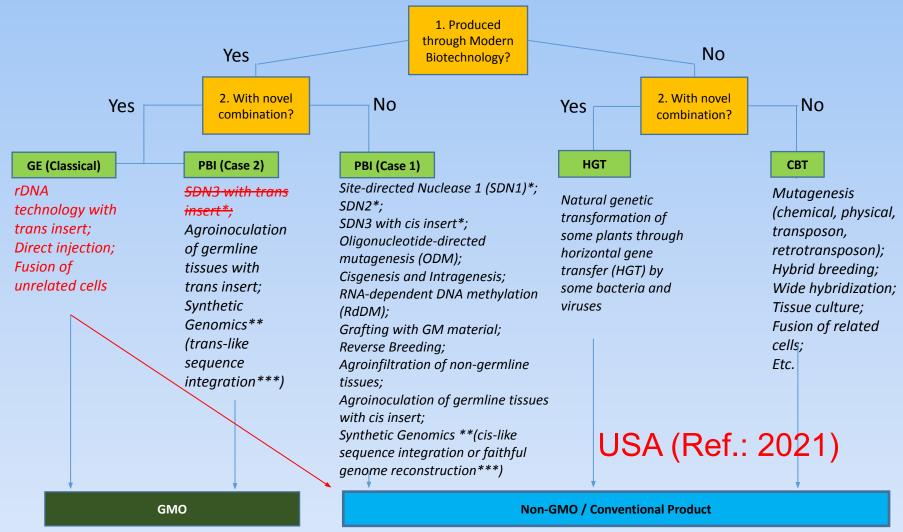


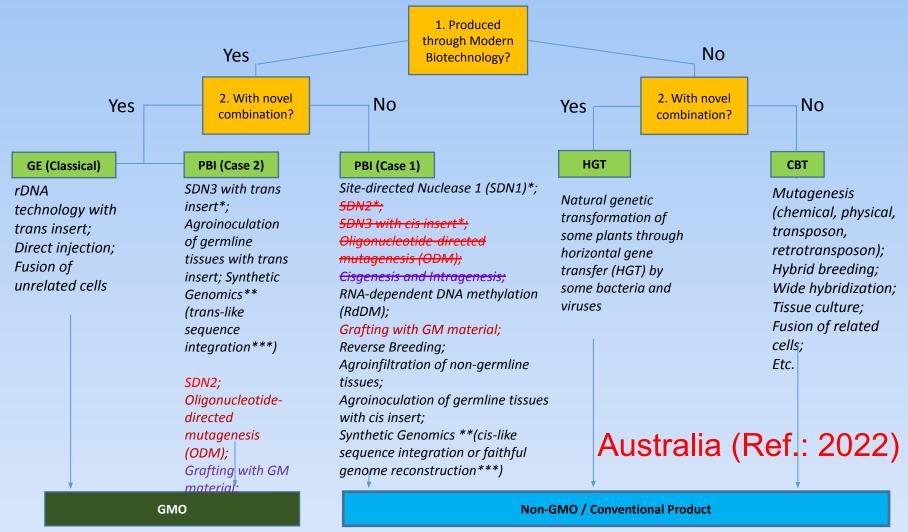


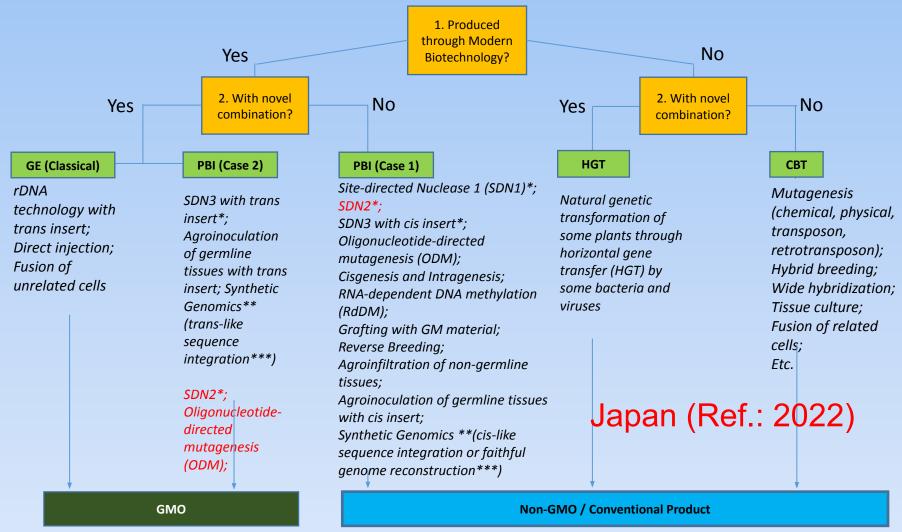


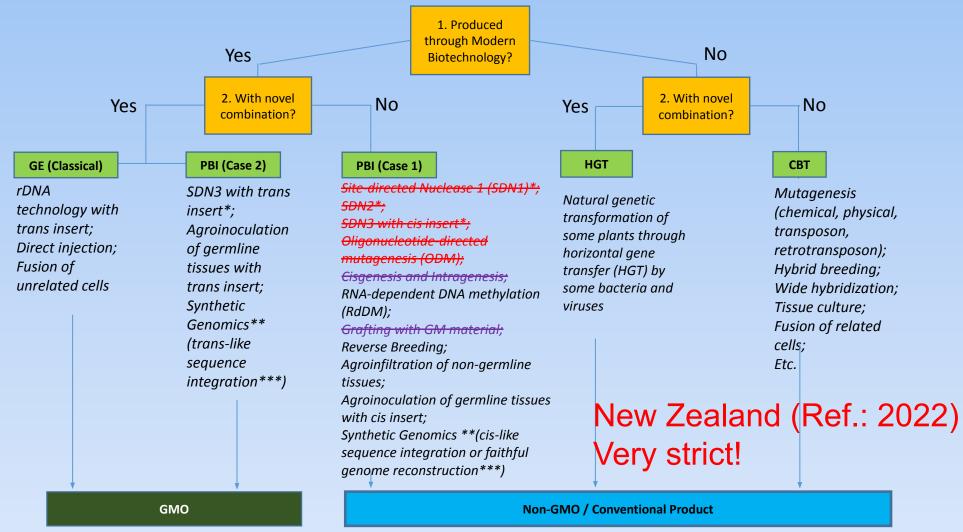


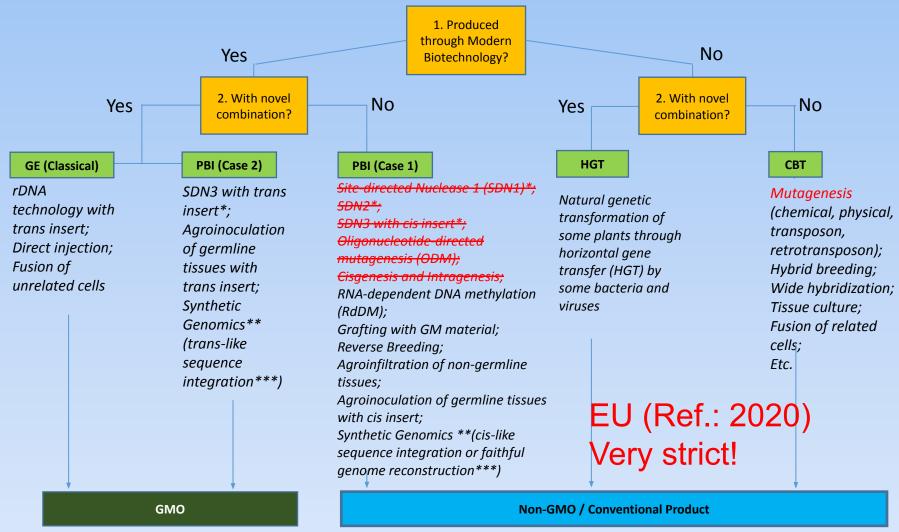
Yes (PNT or GMO) or No (non-GMO)











Key Take-aways

- There are different new technologies available to the modern-day breeders and one of them is Plant Breeding Innovations (PBIs)
- Genome editing is the most popular among the PBIs
- Enabling policies are important for new technologies to have an impact on agriculture
- There are different perspectives globally on how PBIs are assessed (EU and New Zealand are very strict!).













Thank you!













Some New Technologies for Plant Breeding



- Advanced genome/transcriptome sequencing tools (e.g., NGS systems like Illumina, Pacbio, Nanopore)
- Advanced data analytical tools (e.g., bioinformatics, AI)
- 3. Advanced DNA synthesis technologies (up to genome level)
- 4. Plant Breeding Innovations including Precision Breeding



Crops with Whole Genome Sequence information

Name Details

Rice Rice, Os-Nipponbare-Reference-IRGSP-1.0,

Corn, Zm-F7-REFERENCE-TUM-1.0, Zea mays subsp. mays (maize)

Coconut coconut, UPLB_dcnu_1.0, Cocos nucifera (coconut palm)

Coconut coconut, ASM812446v1

Mango Mango, NCBI genome data viewer

Mango Mango, CATAS Mindica 2.1, Mangifera indica (mango), Cultivar: Alphonso

Garlic GARLIC; Allium sativum isolate Ershuizao, whole genome shotgun sequencing proje

Onion ONION; allium cepa DHCU066619 genome sequence

Eggplant Egplant; SME r2.5.1

Tomato Tomato, SL3.0, Solanum lycopersicum (tomato), Cultivar: Heinz 1706

Cotton Cotton, Gossypium hirsutum (cotton), (Mexican cotton)

Rubber tree, ASM165405v1, Hevea brasiliensis (rubber tree), Cultivar: reyan7-33-97

Cassava, M.esculenta_v8, Manihot esculenta (cassava)

Cassava, hifiasm152_l3.hic.hap2.p_ctg, Manihot esculenta (cassava), Cultivar: African

Sweet potato Sweet potato, ipoBat4,Ipomoea batatas (sweet potato), Cultivar: Taizhong6
Potato Potato, SolTub 3.0, Solanum tuberosum (potato), Cultivar: DM 1-3 516 R44

Yam Yam, Dalata_v2, Dioscorea alata (monocots), Cultivar: TDa95/00328, (Purple Yam or Ube) Cacao Cacao, Criollo_cocoa_genome_V2, Theobroma cacao (cacao), Cultivar: B97-61/B2

Papaya Papaya, Papaya1.0, Carica papaya (papaya), Cultivar: SunUp

Banana Banana, ASM31385v2, Musa acuminata subsp. malaccensis (wild Malaysian banana),

Strain: Doubled-haploid Pahang (DH-Pahang)

Abaca upcoming